Genomeditech (Shanghai) Co.,Ltd.
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Product Sheet

H_IL12RB1 HEK-293 Cell Line

Catalog number: GM-C43642

Version 3.3.1.251114

Description	H_IL12RB1 HEK-293 Cell Line is a clonal stable HEK-293 cell line that constitutively expresses the human IL12RB1 gene, constructed using lentiviral technology.
Quantity	5E6 Cells per vial,1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt

Target	Human_IL12RB1
Gene ID/Uniprot ID	P42701-1
Host Cell	HEK-293

Recovery Medium	DMEM+10% FBS+1% P.S
Growth medium	DMEM+10% FBS+1% P.S+125 μg/mL Hygromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO_2

Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

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Materials

Reagent	Manufacturer/Catalogue No.
DMEM	Gibco/C11995500BT
Fetal Bovine Serum	ExCell/FSP500
Pen/Strep	Thermo/15140-122
Hygromycin	Genomeditech/GM-040403
Anti-Human CD212/IL12RB1 Antibody (148C09)	antibodysystem/FHE37110

Figures

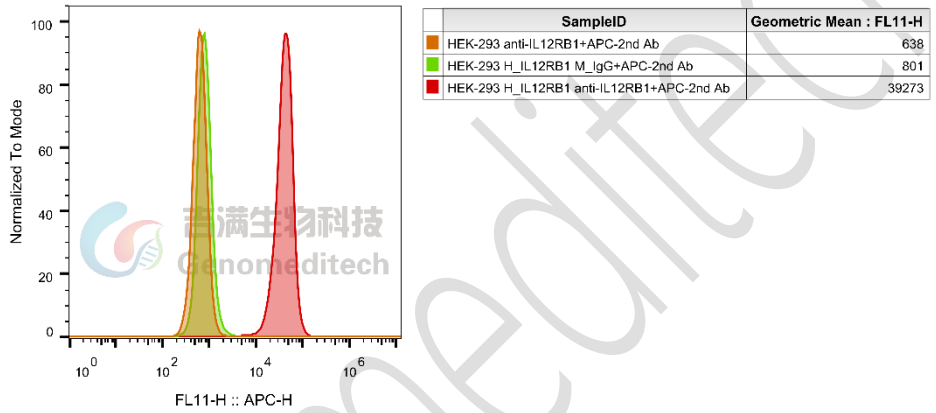

Figure 1 | H_IL12RB1 HEK-293 Cell Line (Cat. GM-C43642) was determined by flow cytometry using Anti-Human CD212/IL12RB1 Antibody (148C09) (AntibodySystem/FHE37110).

Cell Recovery

Recovery Medium: DMEM+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).

b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.

c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.

d) Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.

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e) Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.

b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.

c) Aliquot 1 mL into each vial.

d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: DMEM+10% FBS+1% P.S+125 μg/mL Hygromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

a) Subculturing is necessary when the cell density reaches 80%. It is recommended to perform subculturing at a ratio of 1:3 to 1:4 every 2-3 days. Ensure that the density does not exceed 80%, as overcrowding can lead to reduced viability due to compression.

b) Remove and discard culture medium.

c) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.

d) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 30 to 60 seconds at 37°C).

e) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.

f) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.

g) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.

h) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:3 - 1:4 is recommended

Medium Renewal: Every 2 to 3 days

Notes

a) Upon initial thawing, a higher number of dead cells is observed, which is a normal phenomenon. Significant improvement is seen after adaptation. Once the cells reach a stable state, the number of dead cells decreases after subculturing and the cell growth rate becomes stable.

b) Ensure that the cell density does not exceed 80%, as overcrowding may lead to reduced viability due to compression.

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Sequence

IL12RB1 P42701-1

MEPLVTWVVPLLFLFLLSRQGAACRTSECCFQDPPYPDADSGSASGPRDLRCYRISSDRYECSWQYEGPTAG
VSHFLRCCLSSGRCCYFAAGSATRLQFSDQAGVSVLYTVTLWVESWARNQTEKSPEVTLQLYNSVKYEPPLG
DIKVSKLAGQLRMEWETPDNQVGAEVQFRHRTPSSPWKLGDCGPQDDDTESCLCPLEMNVAQEFQLRRRQL
GSQGSSWSKWSSPVCVPPENPPQPQVRFSVEQLGQDGRRRLTLKEQPTQLELPEGCQGLAPGTEVTYRLQLH
MLSCPCKAKATRTLHLGKMPYLSGAAYNVAVISSNQFGPGLNQTWHIPADTHTEPVALNISVGTNGTTMYW
PARAQSMTYCIEWQPVGQDGGLATCSLTAPQDPDPAGMATYSWSRESGAMGQEKCYYITIFASAHPEKLTL
WSTVLSTYHFGGNASAAGTPHHVSVKNHSLDSVSVDWAPSLLSTCPGVLKEYVVRCRDEDSKQVSEHPVQP
TETQVTLSGLRAGVAYTVQVRADTAWLRGVWSQPQRFSIEVQVSDWLIFFASLGSFLSILLVGVLGYLGLNR
AARHLCPPLPTPCASSAIEFPGGKETWQWINPVDFQEEASLQEALVVEMSWDKGERTEPLEKTELPEGAPELA
LDTELSLEDGDRCKAKM

Related Products

IL-23	
H_IL-23 Reporter 293 Cell Line	H_IL-23 Reporter 293 DDX35TM Cell Line
Cynomolgus_IL-23R HEK-293 Cell Line	H_IL-23R HEK-293 Cell Line
H_IL-23R IL12RB1 HEK-293 Cell Line	Membrane bound H_IL-23 CHO-K1 Cell Line
Anti-IL-23R hIgG1 Antibody(5D4)	
Biotinylated Human IL-23A&IL-12B Heterodimer Protein; His-Avi Tag	Cynomolgus IL-23A & Human IL-12B Heterodimer Protein; His Tag
Cynomolgus IL-23A & Mouse IL-12B Heterodimer Protein; His Tag	Cynomolgus IL-23R Protein; hFc Tag
Human IL-23A & Mouse IL-12B Heterodimer Protein; His Tag	Human IL-23A&IL-12B Heterodimer Protein; His Tag
Human IL-23R Protein; hFc Tag	Human IL-23R Protein; His Tag
Mouse IL-23A&IL-12B Heterodimer Protein; His Tag	
TNF:TNFR2:TNFR1	
H_TNFR2 Null Reporter Cell Line	H_TNFR2 Reporter Jurkat Cell Line
H_TNFR2 Reporter V2 Cell Line	Cynomolgus_TNFRSF1B(TNFR2) CHO-K1 Cell Line
H_TNFRSF1B(TNFR2) CHO-K1 Cell Line	H_TNFRSF1B(TNFR2) HEK-293 Cell Line
Membrane Bound H_TNFα CHO-K1 Cell Line	Membrane Bound H_TNFα(cleavage-resistant) CHO-K1 Cell Line
Anti-H_TNFRSF1B(TNFR2) hIgG1 Antibody(UC2.3.8)	Anti-TNFR1 hIgG1 Antibody(Atrosab)
Human TNF alpha Protein; His Tag	Human TNFR1 Protein; hFc Tag
Human TNFR2 Protein; hFc Tag	
TL1A:DR3(TNFRSF25)	
H_TNFRSF25(DR3) Reporter Jurkat Cell Line	H_TNFSF15(TL1A) Reporter Cell Line
Mouse_TNFRSF25(DR3) Reporter Jurkat Cell Line	Cynomolgus_TNFSF15(TL1A) HEK-293 Cell Line
H_TNFRSF25(DR3) CHO-K1 Cell Line	H_TNFRSF25(DR3) HEK-293 Cell Line
H_TNFSF15(TL1A) CHO-K1 Cell Line	H_TNFSF15(TL1A) HEK-293 Cell Line
Mouse_TNFSF15(TL1A) HEK-293 Cell Line	

Anti-H_TNFSF15(TL1A) hIgG1 Antibody(PF-06480605)	Anti-H_TNFSF15(TL1A) hIgG1 Antibody(Tulisokibart、PRA-023)
Anti-H_TNFSF15(TL1A) hIgG4 Antibody	Anti-TL1A hIgG1 Reference Antibody (Duvbio)
Anti-TL1A hIgG1 Reference Antibody (Tulbio)	Anti-TNFRSF25(DR3) hIgG1 Antibody(PTX-35)
Biotinylated Cynomolgus TL1A Protein; His-Avi Tag	Biotinylated Human TL1A Protein; His-Avi Tag
Cynomolgus TL1A Protein; His Tag	Human TL1A Protein; His Tag

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